

How to combat bacteriophages that disrupt food fermentations

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Moved
in 1994
To Cork

The Netherlands



Ireland



Milk-to-cheese conversion using (traditional) biotechnology



Dairy fermentations using
Lactic Acid Bacteria or LAB
(*Lactococcus lactis/cremoris*)

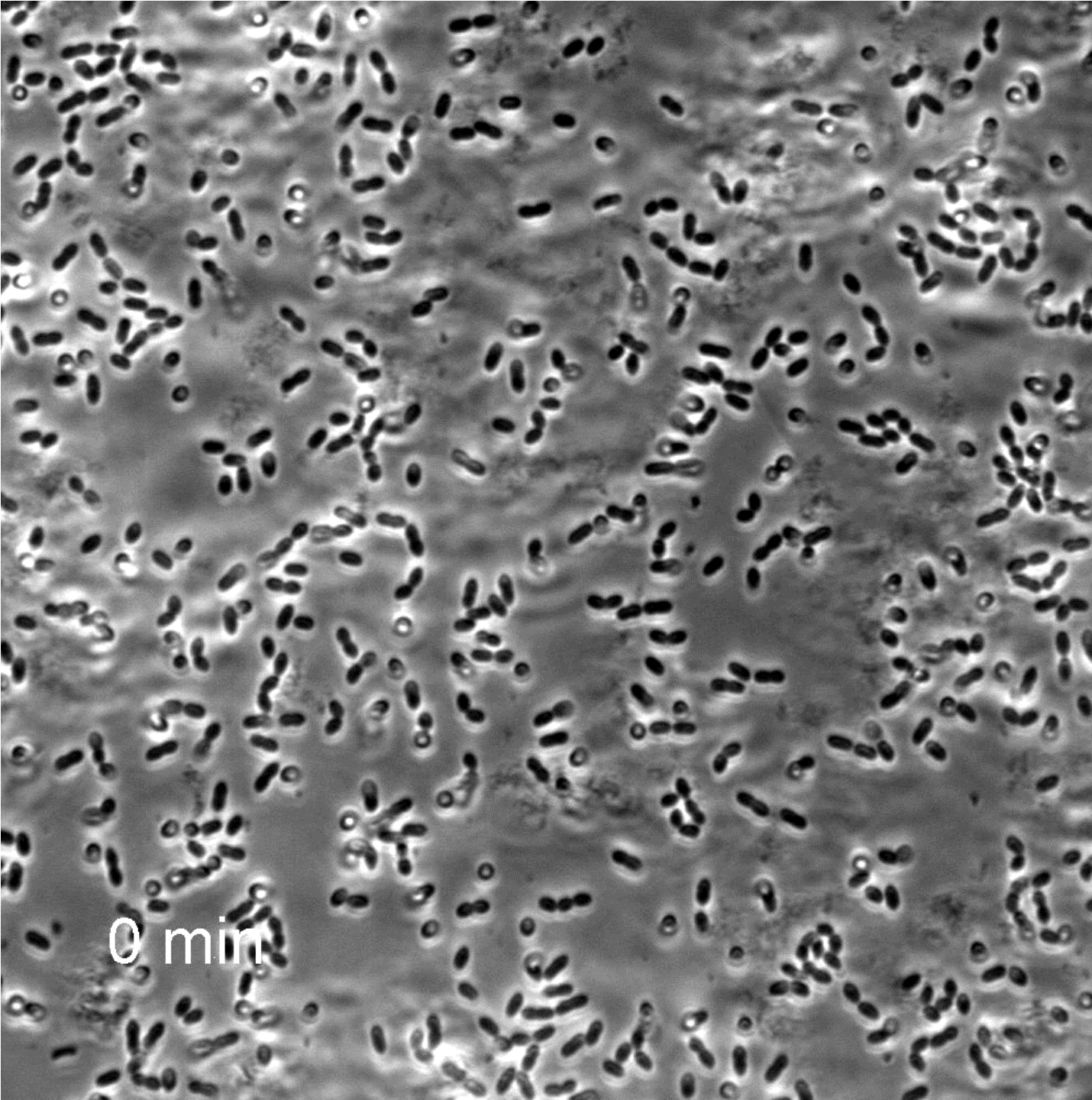
Annual cheese production:

~23 million metric
Tonnes (2025 projection)

Value: 55 billion dollars

This milk conversion process involves approx. two
quintillion (2,000,000,000,000,000,000) bacteria!

Ideal for phage infections, causing
problems in the fermentation process



L. lactis plus
bacteriophage

Courtesy:
James Murphy (UCC)
& Târn Mignot (CNRS,
Marseille)

What can we do to prevent/minimize damage caused by phage attack of starter cultures?

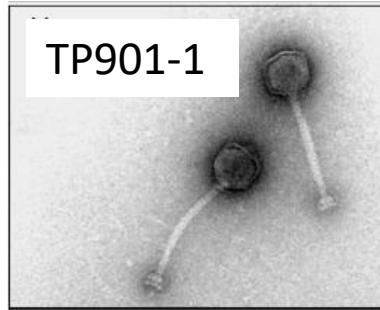
Lots of research has been done:

- A given phage will only **attack a few strains** of *L. lactis*
- Milk is **not sterile** and introduces phages into factory
- Once in the factory, **impossible** to eradicate

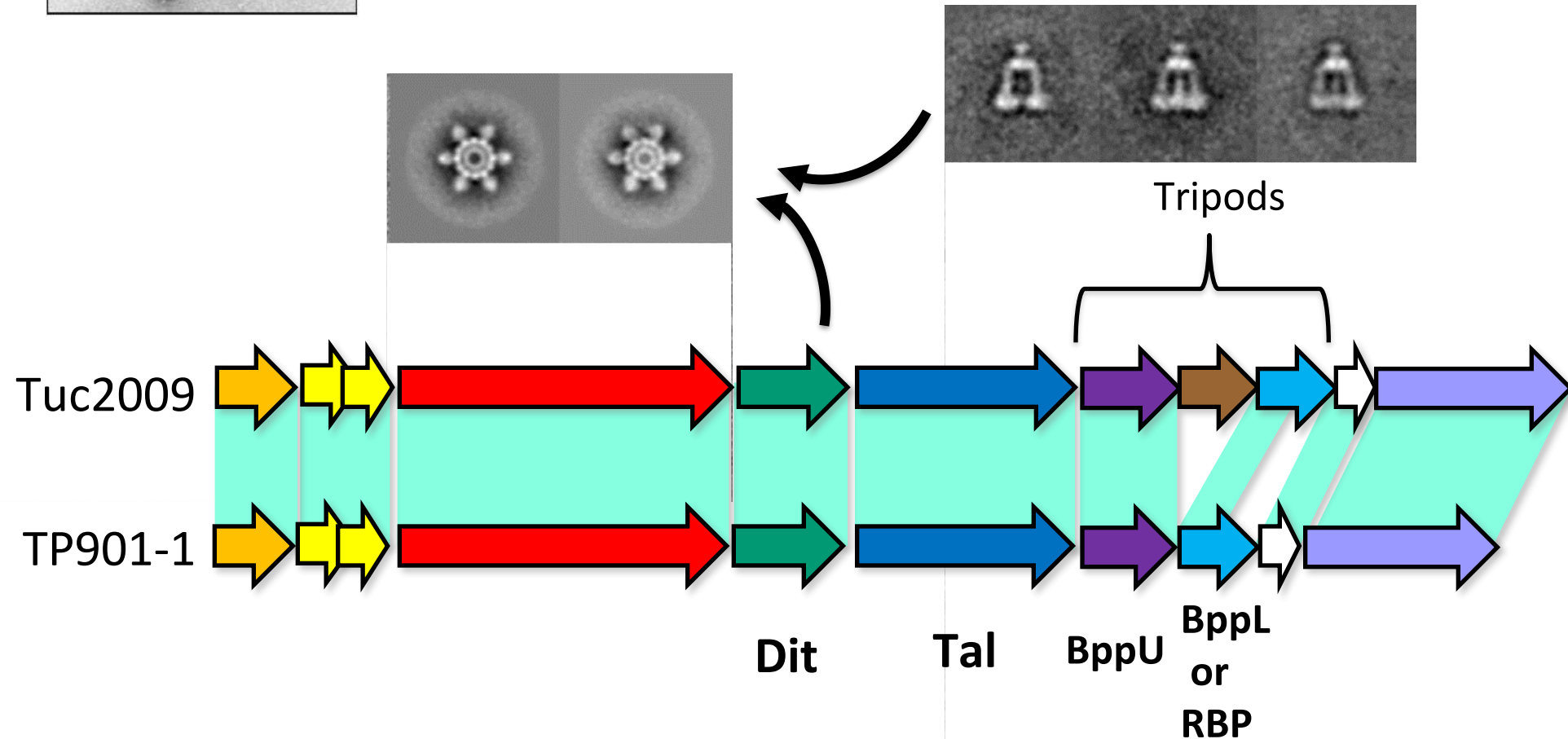
Three major questions:

- **How** does a phage recognize a lactococcal host?
- **What** does it recognize?
- **Can** we mobilize indigenous phage immunity?

Tuc2009 & TP901-1 as lactococcal model phages



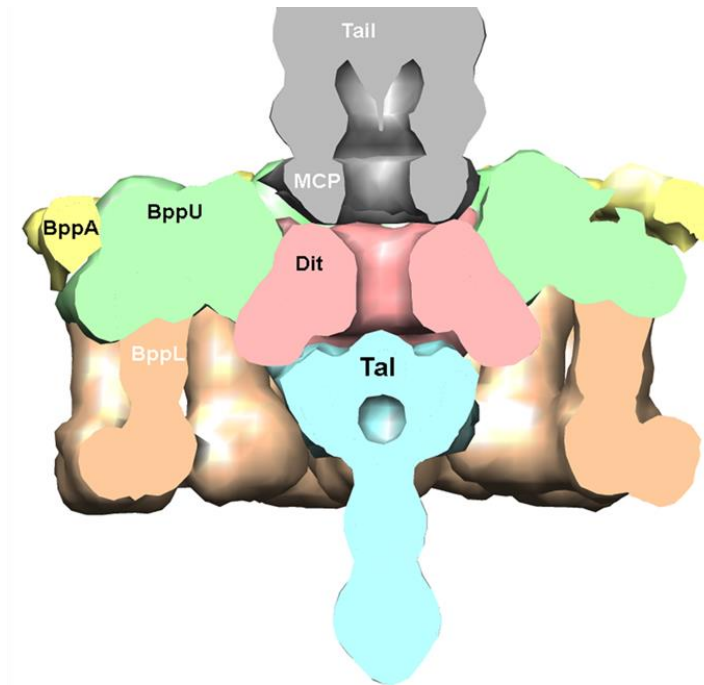
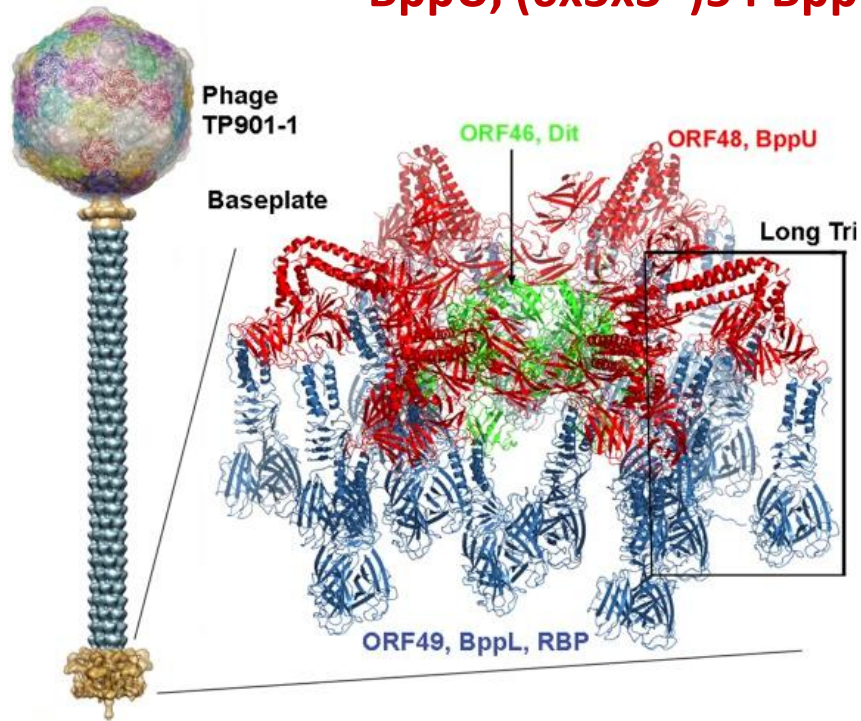
- Infect distinct strains though highly homologous



Structure of the phage TP901-1 1.8 MDa baseplate suggests an alternative host adhesion mechanism

David Veessler^{a,b,1,2}, Silvia Spinelli^{a,b}, Jennifer Mahony^c, Julie Lichère^{a,b}, Stéphanie Blangy^{a,b}, Gérard Bricogne^d, Pierre Legrand^e, Miguel Ortiz-Lombardia^{a,b}, Valérie Campanacci^{a,b,3}, Douwe van Sinderen^{c,f}, and Christian Cambillau^{a,b,1}

Heteromultimeric baseplate complex: 6 Dit, (6x3=)18 BppU, (6x3x3=)54 BppL (RBP), ([6x2=]12 BbbA)

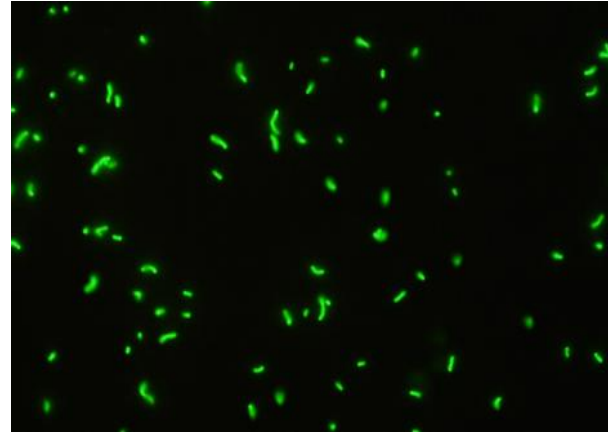


Veesler et al., PNAS, 2012; Legrand et al., mBio, 2016;
Hayes et al., Mol Microbiol, 2018; Hayes et al., Viruses, 2019

L. cremoris UC509.9

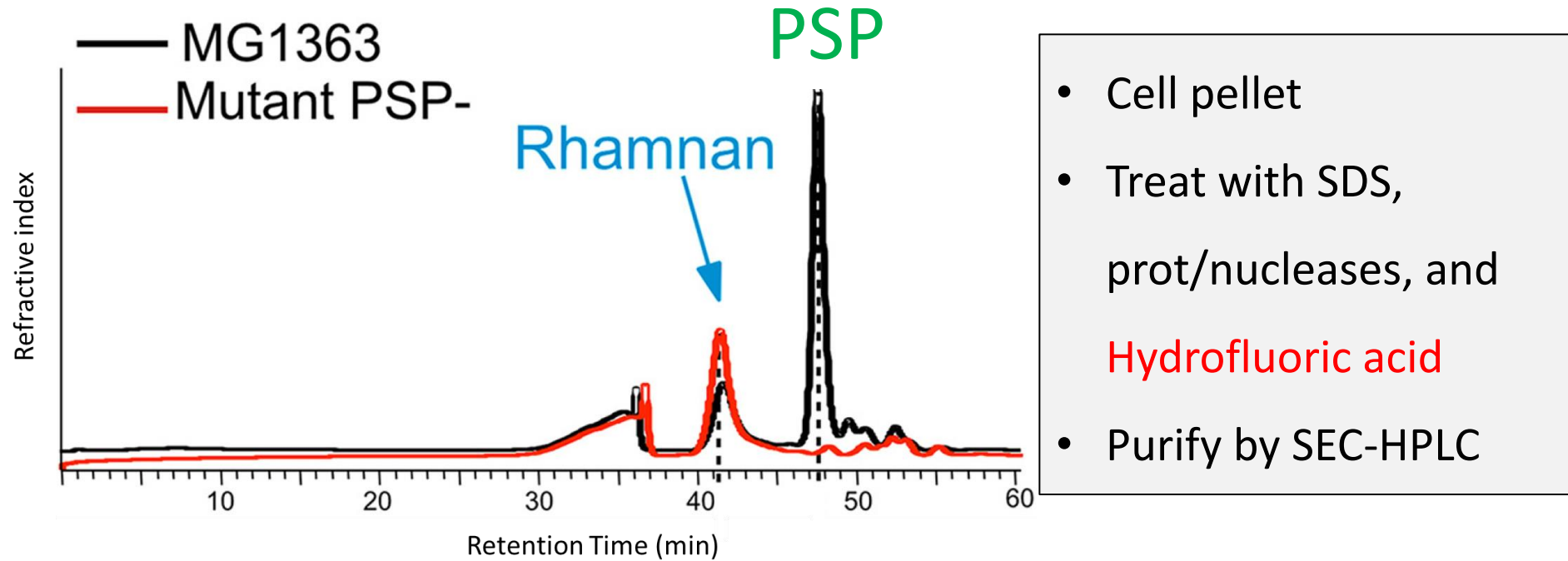
L. cremoris 3107

GFP-labelled
Tripods Tuc2009



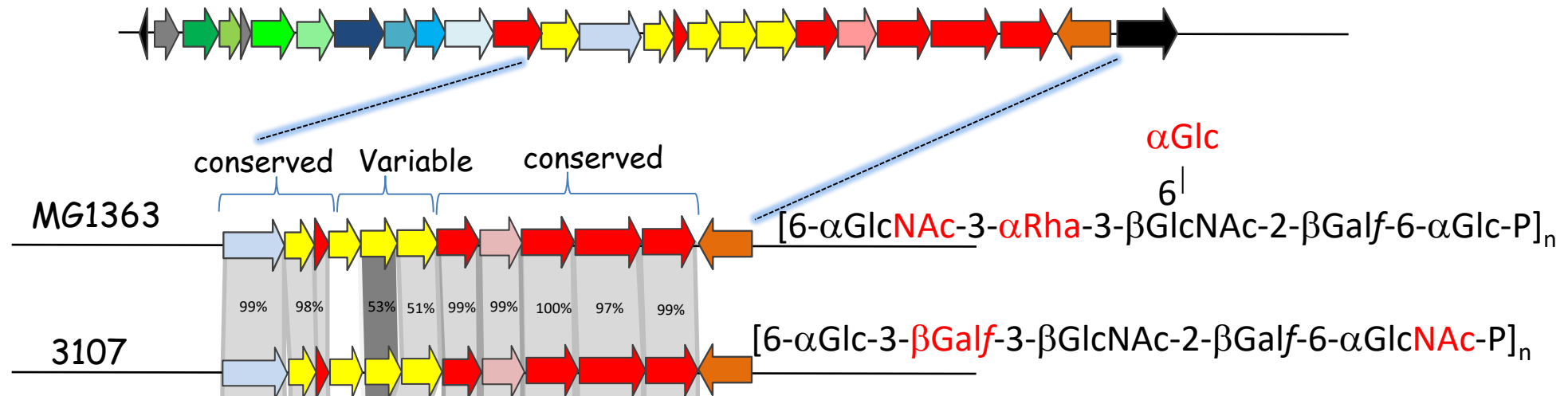
What does the phage recognize on the host???

The lactococcal cell envelope contains a cell wall polysaccharide (CWPS):
which consists of a conserved rhamnan plus a variable pellicle or PSP



Mutations in the *cwps* cluster of *L. lactis* MG1363 cause a PSP-negative phenotype and phage insensitivity

Genetic diversity among *cwps* cluster and PSP structure

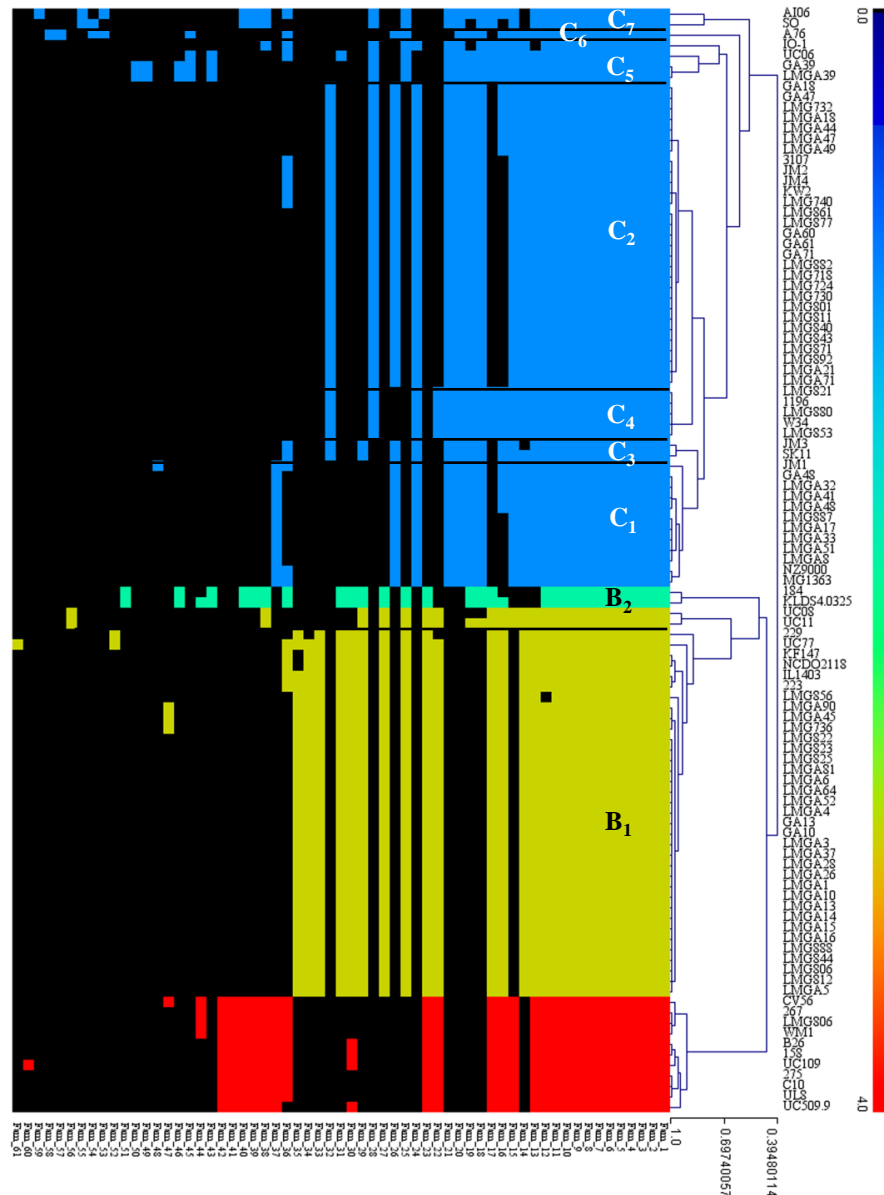


Gene swapping of the variable region leads to PSP inversion (NMR surface scanning) and to host range swapping of TP901-1

Ainsworth et al., mBio, 2014

Mahony et al., AEM, 2015

L. Lactis genome sequencing reveals four cwps genotypes



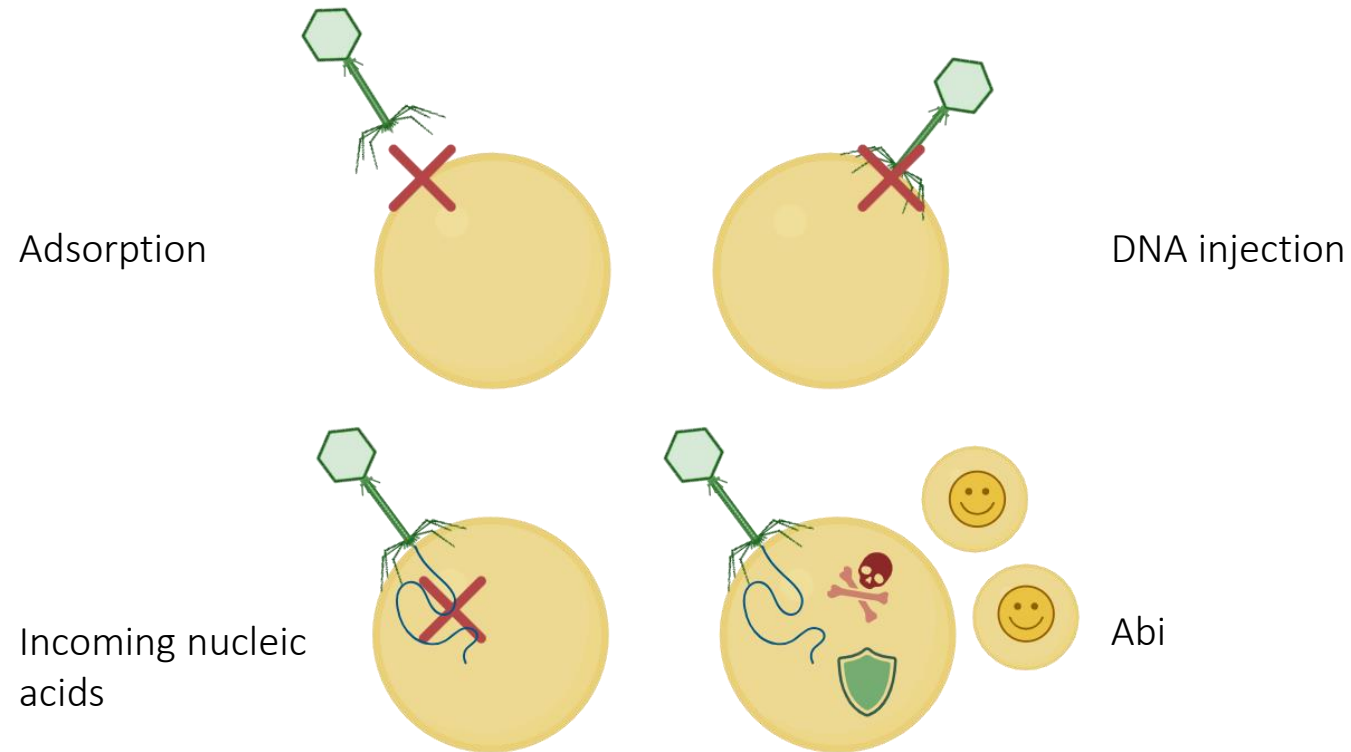
C Structures of representatives of each *cwps* genotype have been determined

D RBP variation is correlated to the CWPS type of corresponding host (**rational starter culture design**)

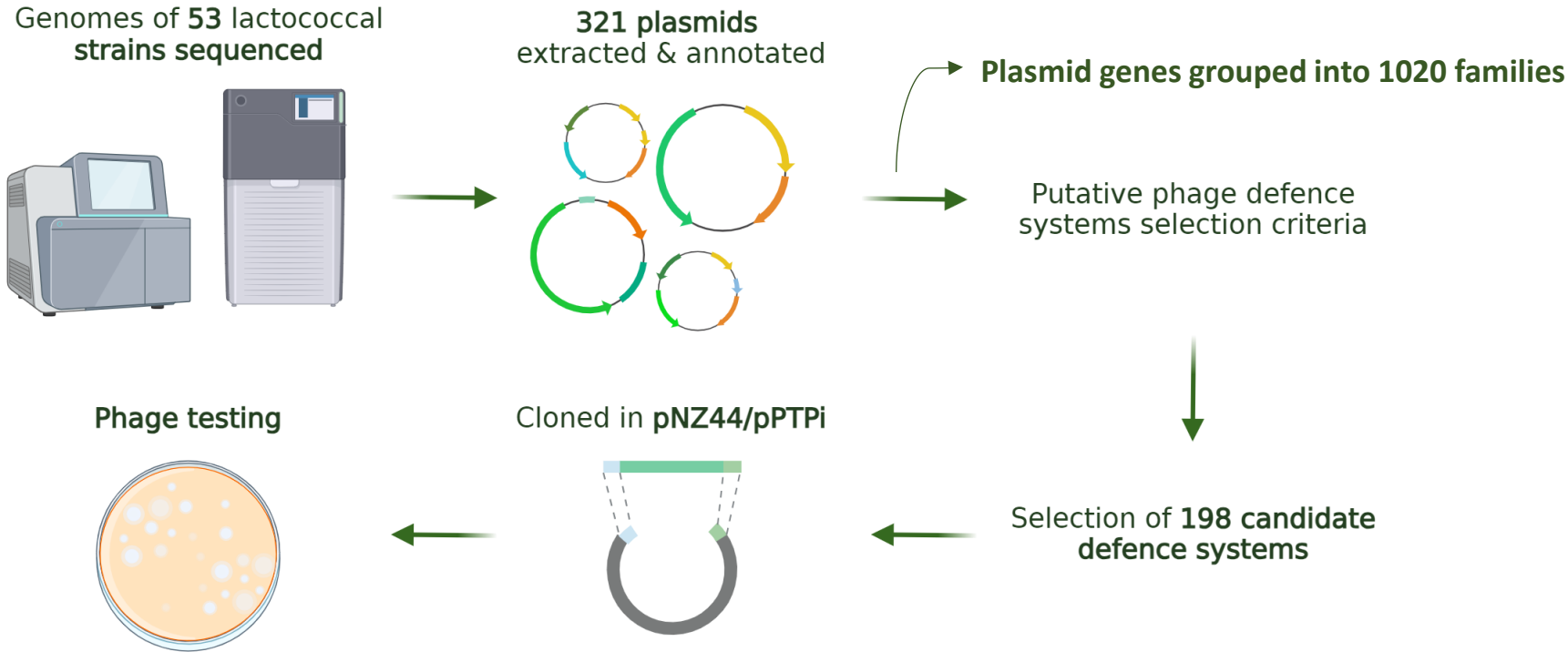


Bacterial immunity

- Multiple and diverse bacterial phage defence systems
- Abortive infection (Abi) - one of the most common system in *Lactococcus*



Identification of novel antiphage systems



Nucleic Acids Research, 2024, **52**, 9760–9776
<https://doi.org/10.1093/nar/gkaf671>
Advance access publication date: 9 August 2024
Molecular Biology



Discovery of antiphage systems in the lactococcal plasmidome

Andriana Grafakou^{1,†}, Cas Mosterd^{1,†}, Matthias H. Beck¹, Philip Kelleher¹, Brian McDonnell¹, Paul P. de Waal², Irma M.H. van Rijswijck², Noël N.M.E. van Peij², Christian Cambillau^{1,3}, Jennifer Mahony¹ and Douwe van Sinderen^{1,*}



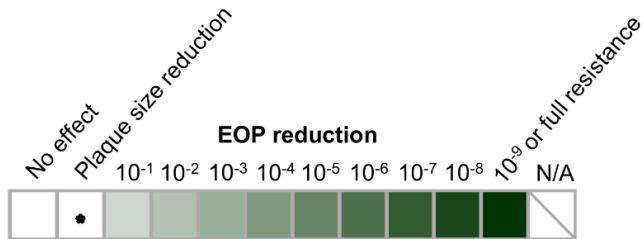
Antiphage activity spectrum

- 20 functional antiphage systems identified
- All except two effective against most frequently encountered phages
- Many systems on conjugative plasmids



Lactococcal strain phage

- *L. cremoris* NZ9000
- *L. lactis* IL1403
- *L. cremoris* 3107



N/A for either cloned in pPTPi or non-transformable/inconsistent in 3107

New antiphage systems

Systems new in
Lactococcus

Known lactococcal Abi

	Skunavirus										Ceduvirus	P335 group			Teubervirus			
	sk1	712	P2	jj50	bIL66	bIL70	P008	P113G	340	66901	62601	C2	P335	TP901-1	LC3	Dub35A	63301	P087
Rhea (pNZ44)	*	*		*	*		*			*	*	*	*					
Aristaios (pNZ44)																		
Kamadhenu (pNZ44)																		
Fliodhais (pPTPi)												*						
Audmula (pNZ44)																		
Rugutis (pNZ44)					*	*	*	*	*			*						
Hesat (pPTPi)																		
PARIS (pPTPi)	*	*	*	*								*						
Type I CBASS (pNZ44)												*						
Type II CBASS (pNZ44)																		
Lamassu (pPTPi)																		
Septu (pPTPi)			*															
AbiA (pPTPi)												*						
AbiB-like (pNZ44)	*	*	*	*	*	*	*		*	*								
AbiD-like (pNZ44)	*	*	*	*		*	*		*		*							
AbiF (pNZ44)									*	*		*						
AbiG (pNZ44)			*		*	*	*	*	*									
AbiJ (pNZ44)	*	*	*									*						
AbiP (pNZ44)	*	*				*	*	*										
AbiZ (pNZ44)	*	*	*		*	*	*		*									

- We know how phage recognizes host
- We can predict phage sensitivity of a strain
- Rational starter culture design

- Many new anti-phage systems identified (also in *S. thermophilus* (Kelleher et al., NAR, 2024)
- Allows ‘construction’ of phage immunity into lactococcal strains
- Problem solved!?

Collaborators

Christian Cambillau, Silvia Spinelli, Renaud Vincentelli & Tam Mignot, CNRS, Marseille, France

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Pierre Legrand, St Aubin, France

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Peptidoglycan

